

## ❑ Proteins: why are so important?

❑ From the Greek: **“Being of Primary Importance”**

No better world could have been chosen

**Enzymatic catalysis**

**Transport and Storage**

**Movement**

**Immunitary Defense**

**Transmission of nervous signals**

**Hormon activity**

**Forming tissues**

**Albuminoids**: these materials were identified (1800) in natural processes as the coagulation of egg white by heat, the curdling of milk with acid or the spontaneous clotting of blood

Gerardus Mulder (1802-1880) proposed a molecular form:  $C_{40}H_{62}N_{10}O_{12}$   
+ S

Justus von Liebig(1803-1873):

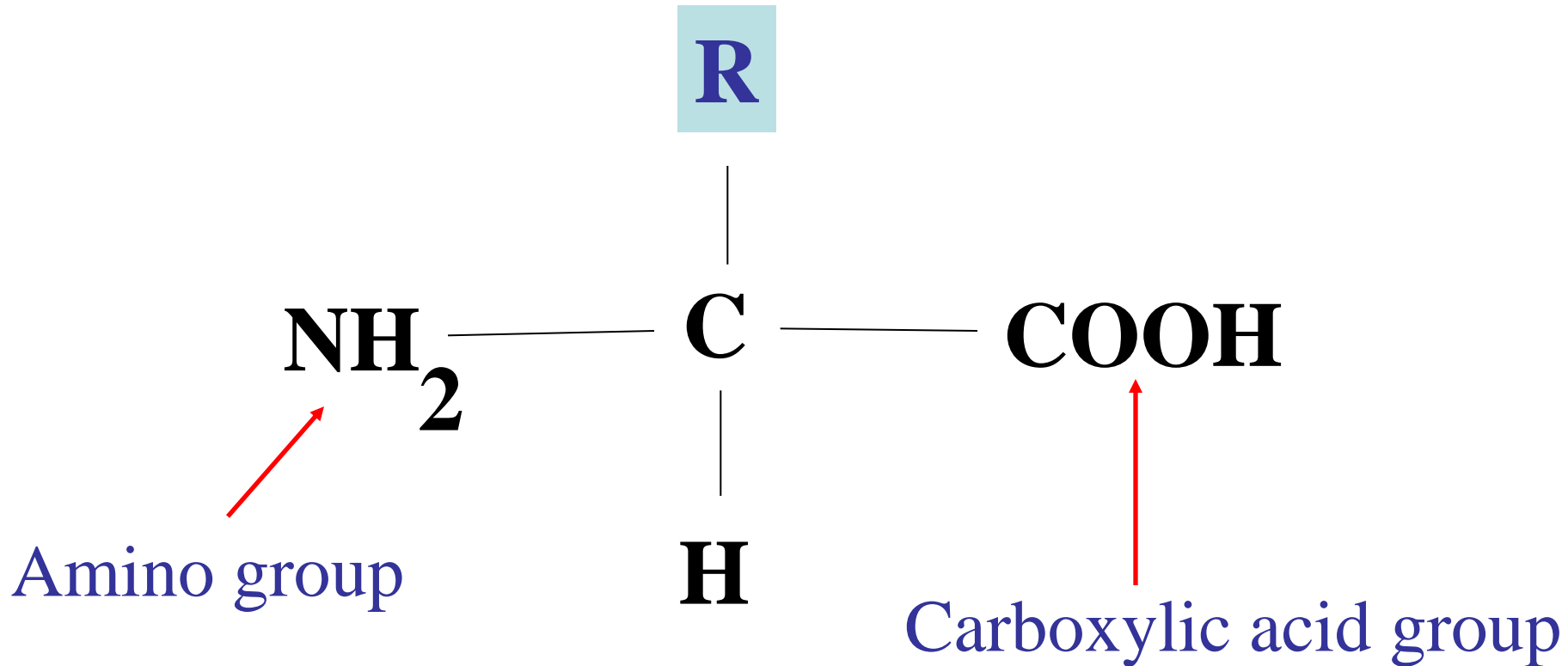
We cannot isolate the particular substance described by Mr. Moulder. And so, it is a source of despair, after so much has been prattled and written about “protein”, to have to say there is no such thing.

Mulder again:  $C_{36}H_{54}N_8O_{12}$

Von Liebig (working with casein) purified to small molecules:

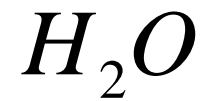
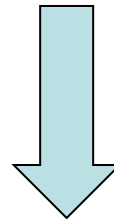
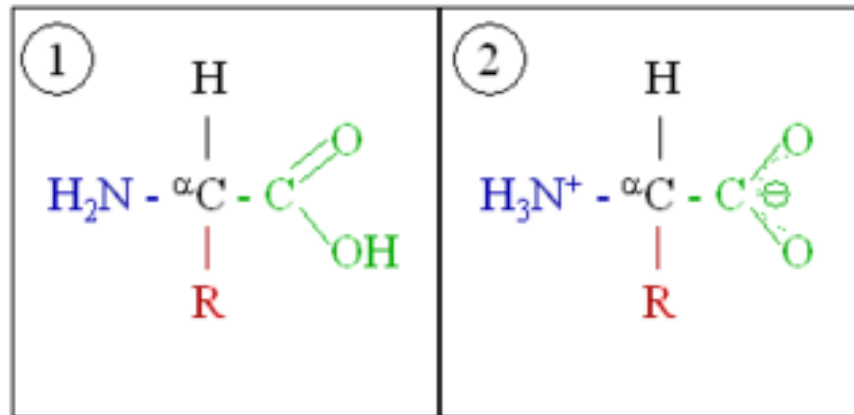
first amino-acids: leucine and tyrosine

# 20 KIND OF AMINO-ACIDS

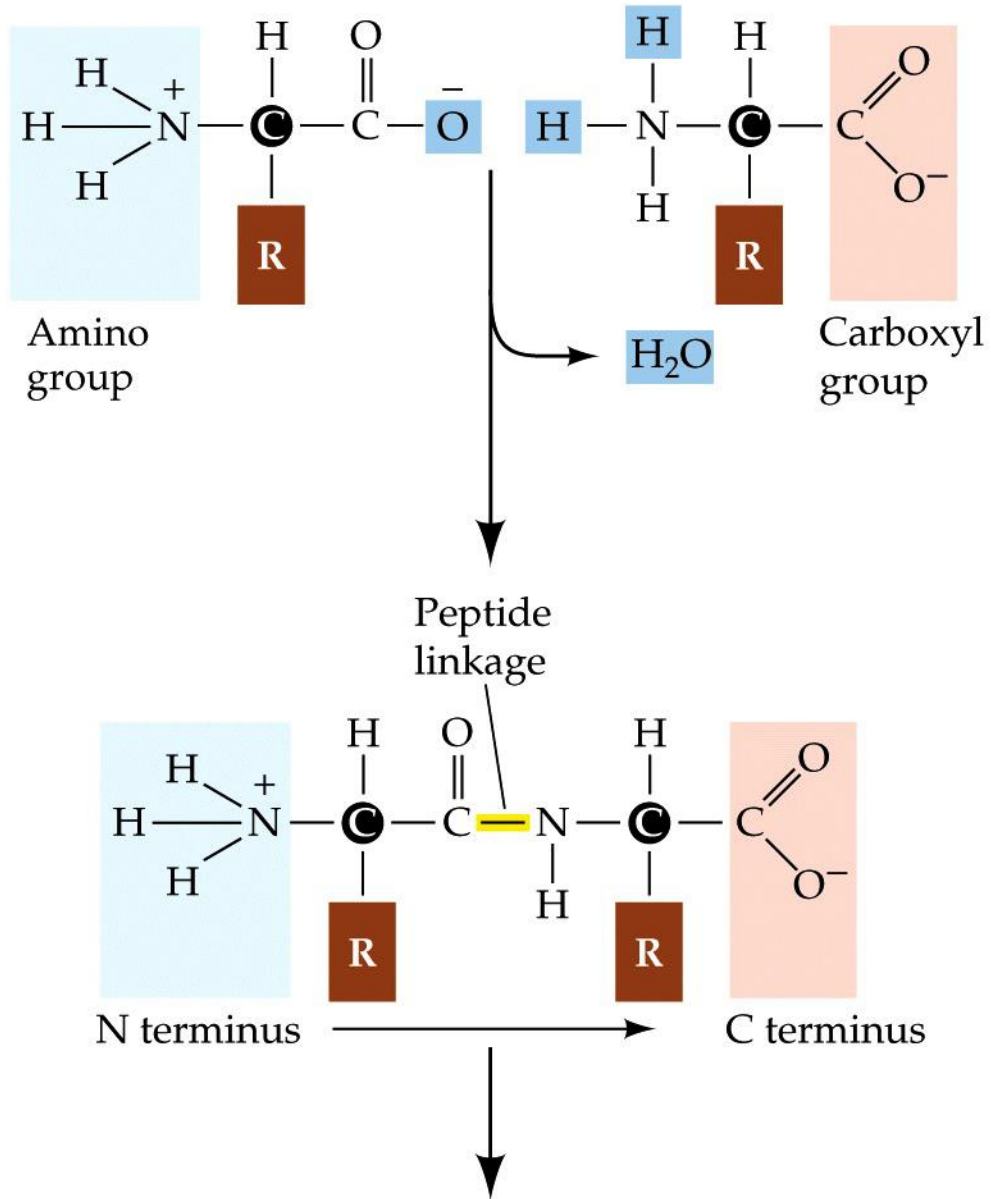
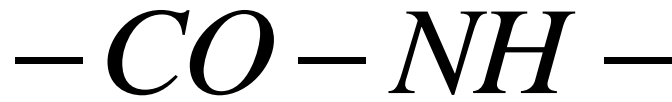




Covalent Bond



Covalent Bond



# Protein Chain

Carbon alpha, Carbon', Nitrogen, .... Hydrogen omitted

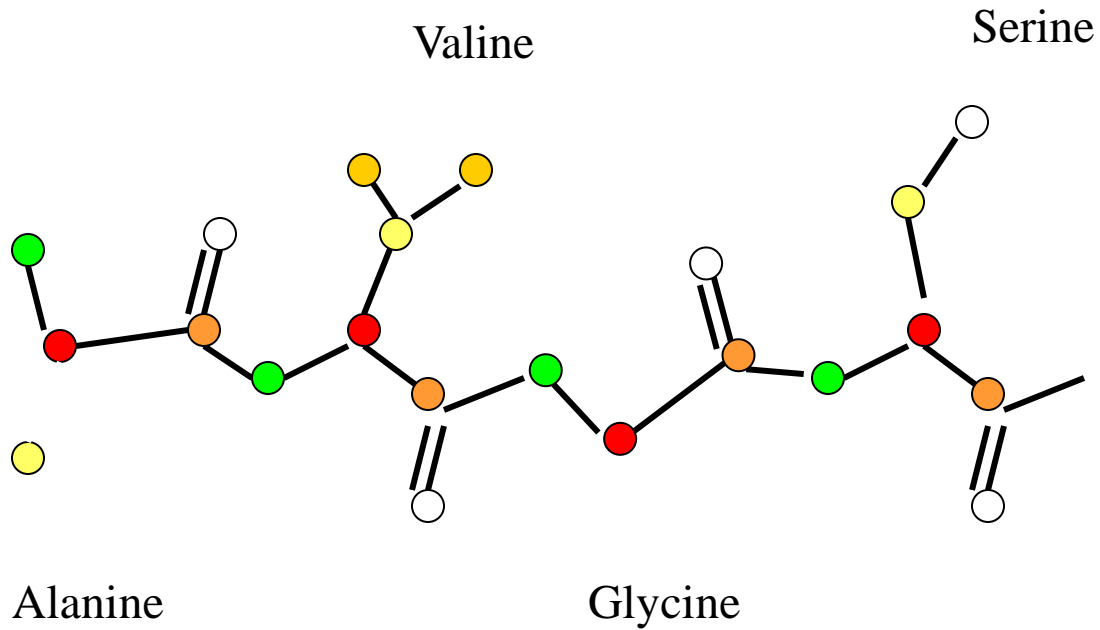


Table 1

## Languages

Each monomer is identified by a symbol

Amino acid names and abbreviations		
Amino acid	Three letter code	Single letter code
Alanine	Ala	A
Arginine	Arg	R
Aspartic acid	Asp	D
Asparagine	Asn	N
Cysteine	Cys	C
Glutamic acid	Glu	E
Glutamine	Gln	Q
Glycine	Gly	G
Histidine	His	H
Isoleucine	Ile	I
Leucine	Leu	L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
Serine	Ser	S
Threonine	Thr	T
Tryptophan	Trp	W
Tyrosine	Tyr	Y
Valine	Val	V

2nd base in codon

1st base in codon

	U	C	A	G	
U	Phe Phe Leu Leu	Ser Ser Ser Ser	Tyr Tyr STOP STOP	Cys Cys STOP Trp	U C A G
C	Leu Leu Leu Leu	Pro Pro Pro Pro	His His Gln Gln	Arg Arg Arg Arg	U C A G
A	Ile Ile Ile Met	Thr Thr Thr Thr	Asn Asn Lys Lys	Ser Ser Arg Arg	U C A G
G	Val Val Val Val	Ala Ala Ala Ala	Asp Asp Glu Glu	Gly Gly Gly Gly	U C A G

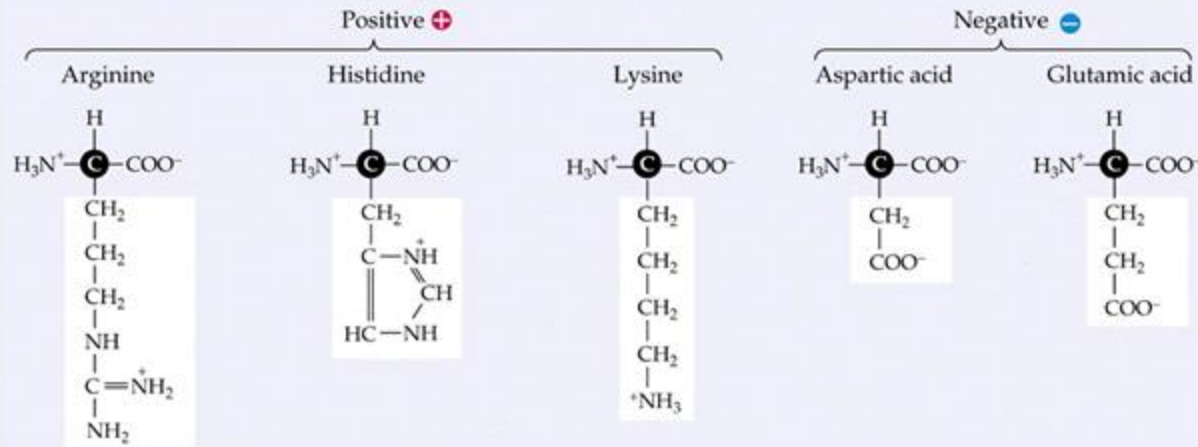
3rd base in codon



# Amino-acidi Carichi

## 3.2 Twenty Amino Acids Found in Proteins

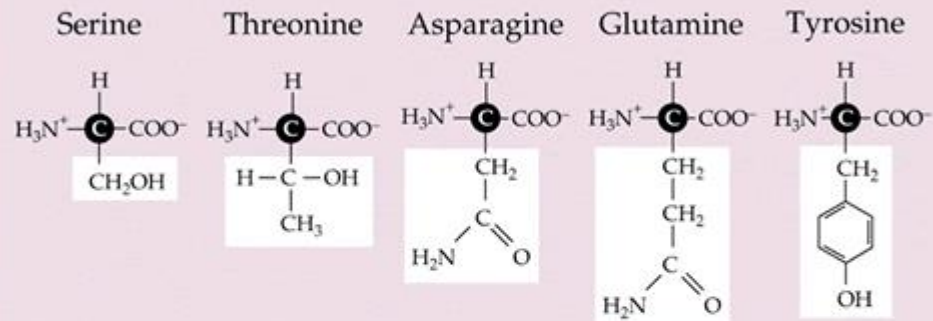
### A. Amino acids with electrically charged side chains



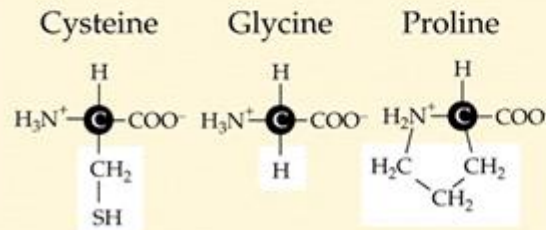
# Amino-acidi Polari (idrofilici)

## 3.2 Twenty Amino Acids Found in Proteins

### B. Amino acids with polar but uncharged side chains



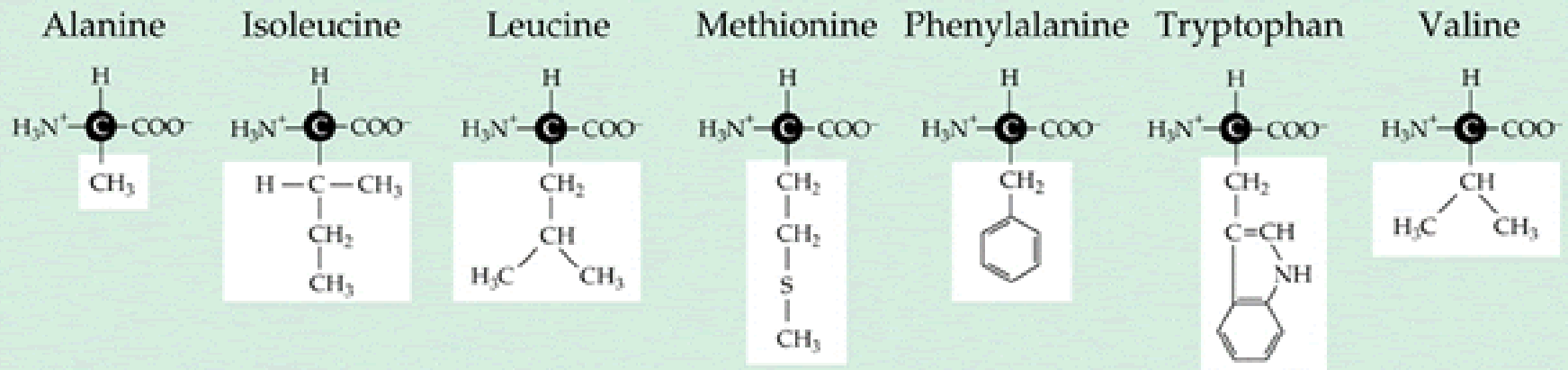
### C. Special cases



# Amino-acidi non polari (idrofobici)

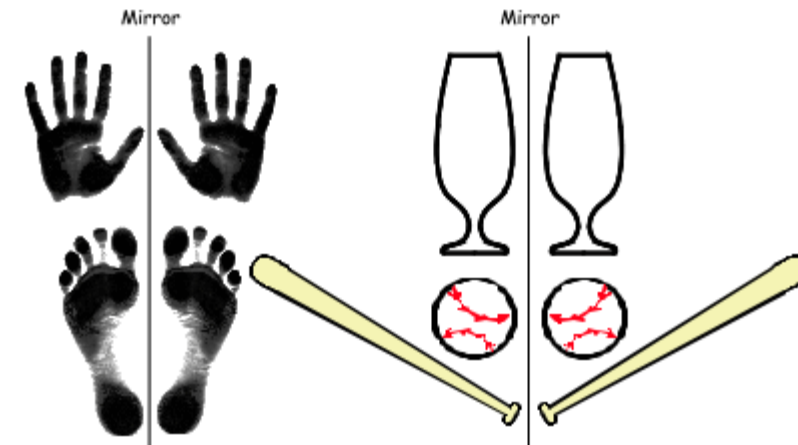
## 3.2 Twenty Amino Acids Found in Proteins

### D. Amino acids with hydrophobic side chains



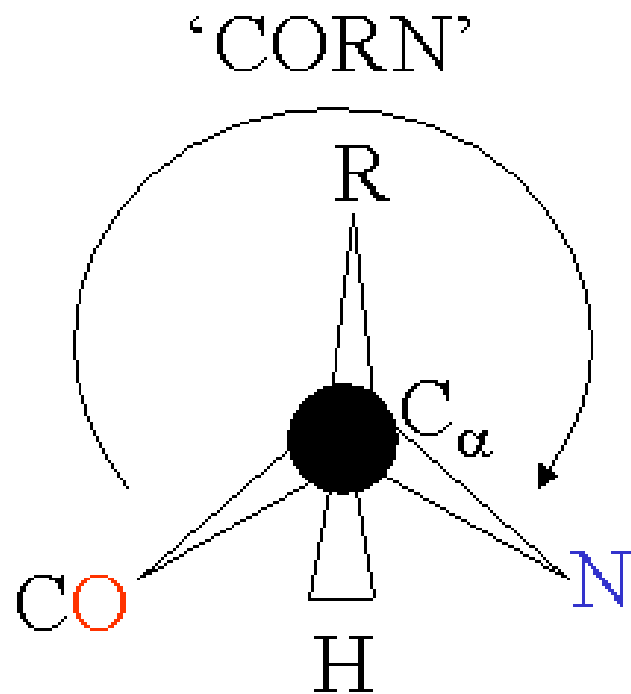
# CHIRALITY

An object that cannot be superimposed on its mirror image is called chiral



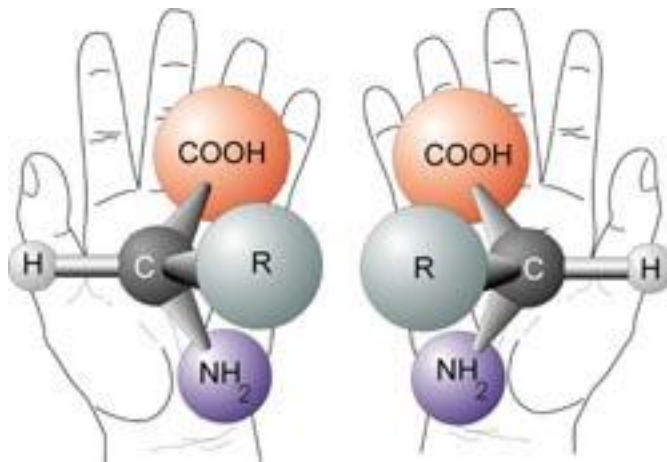
**Chiral objects**  
Nonsuperimposable  
mirror images

**Nonchiral objects**  
Superimposable  
mirror images



Guardando  $C_{\alpha}$  da  $H$

A **chiral molecule** is a type of [molecule](#) that lacks an internal plane of [symmetry](#) and has a non-superimposable [mirror image](#). The feature that is most often the cause of [chirality](#) in molecules is the presence of an [asymmetric carbon atom](#).]



Although most amino acids can exist in both left and right handed forms, Life on Earth is made of left handed amino acids, almost exclusively. No one knows why this is the case. However, Cronin and Pizzarello have shown that some of the amino acids that fall to earth from space are more left than right. Thus, the fact that we are made of L amino acids may be because of amino acids from space.

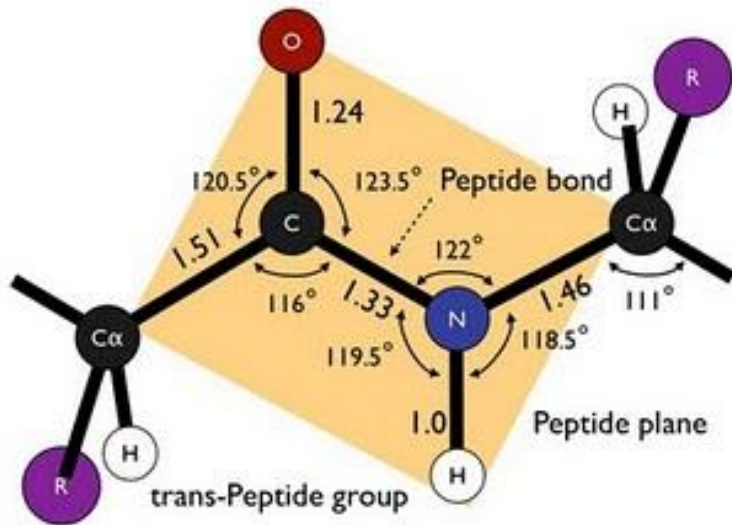
La prima proteina (myoglobina) fu cristallizzata nel 1961.

La selezione di una proteina di N aminoacidi non puo' essere avvenuta per trial and error.

Eta' dell'universo  $5 \cdot 10^{17}$  secondi

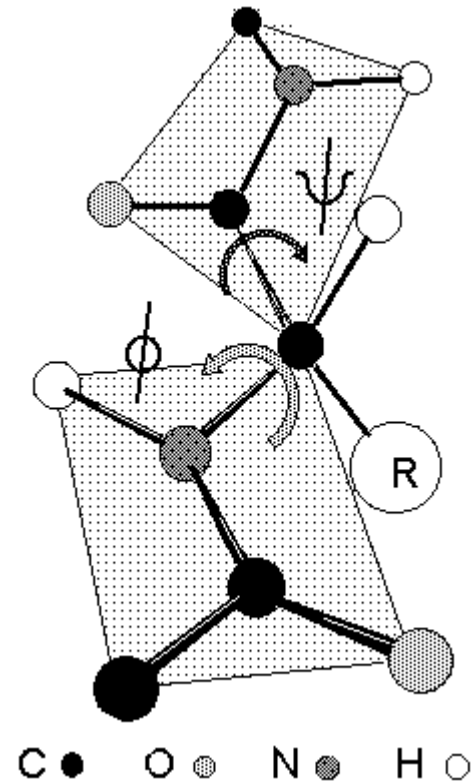
Tempo necessario per esplorare  $20^N$  enorme  $20^{300} = 10^{400}$

Una proteina con nuova funzione puo' risultare dalla fusione di mRNA di sequenze piu' corte, ognuna delle quali selezionata per una funzione piu' semplice.



$C - N = 1.47 \text{ \AA}$   
 $C = N = 1.25 \text{ \AA}$   
 nel legame peptidico:  
 $C - N = 1.32 \text{ \AA}$

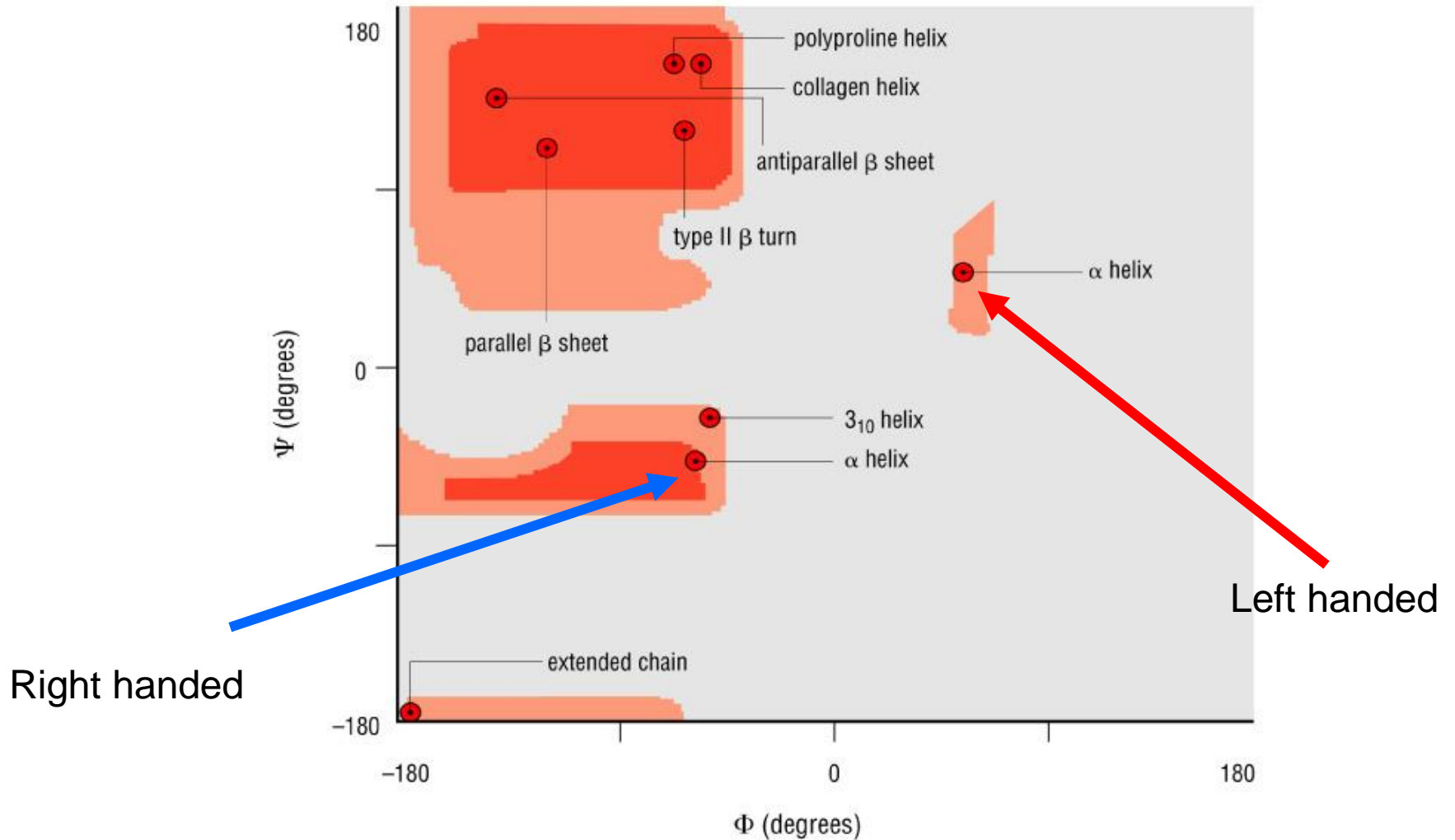
Trans favorito rispetto a Cis  
 $4 \text{ kcal mol}^{-1}$





# Grafico di Ramachandran

From [Protein Structure and Function 2004-2005 Online Update](#)  
by Gregory A Petsko and Dagmar Ringe

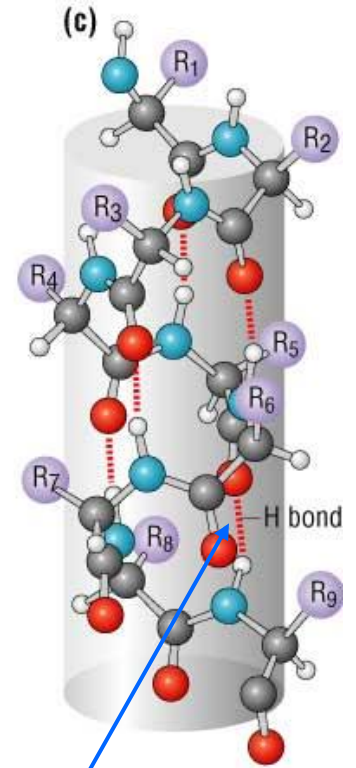
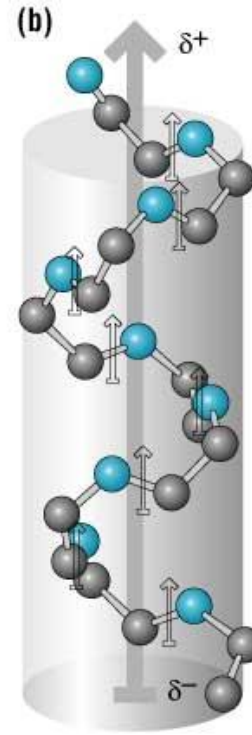
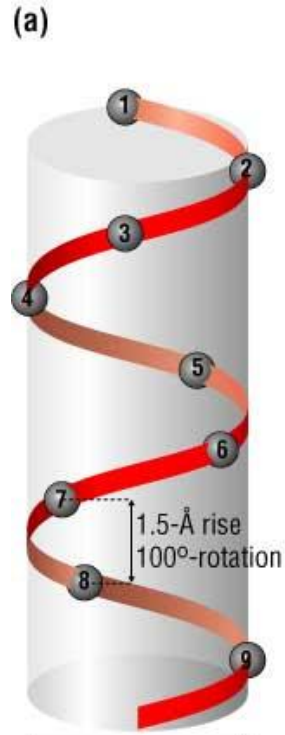


# $\alpha$ helices

$p=1.5 \text{ \AA}$

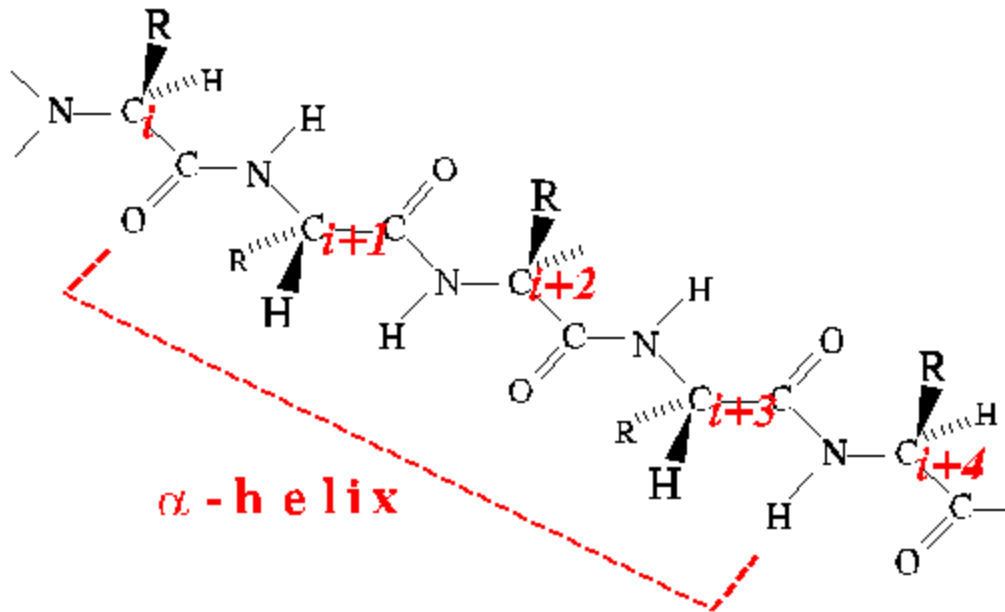
$P/p=3.6$

Hb  $\{i, i+4\}$



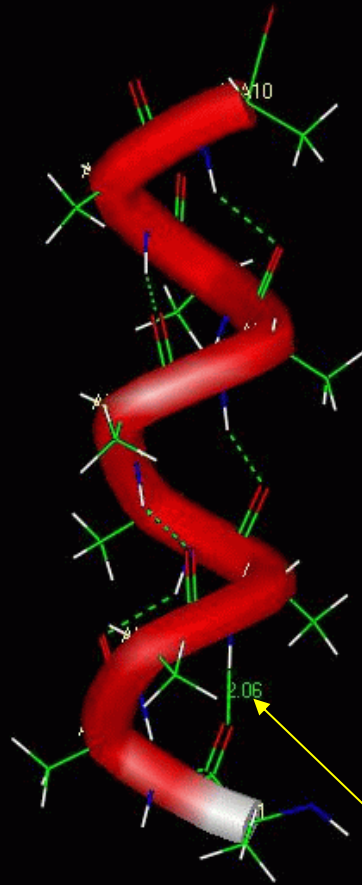
Parallel to the axis

Right-handed



# Elica 3-10

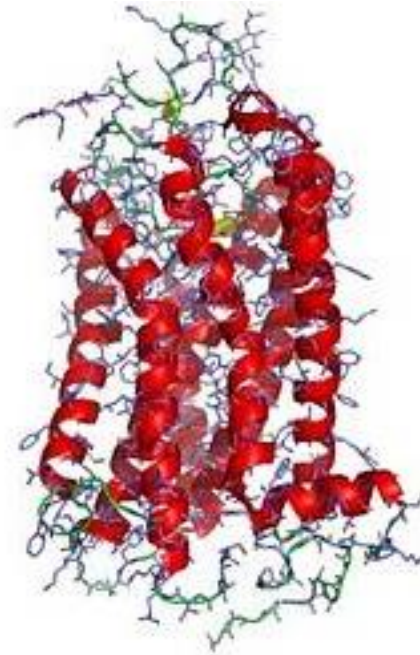
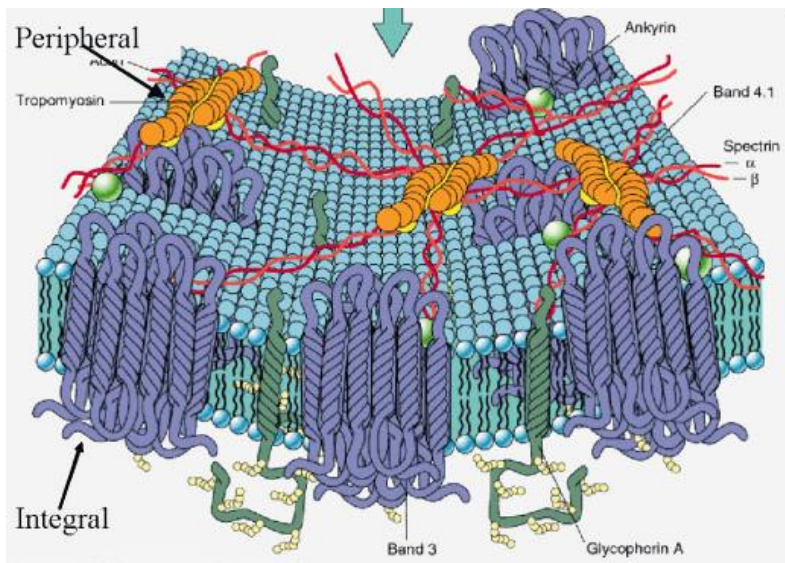
3-10 helix



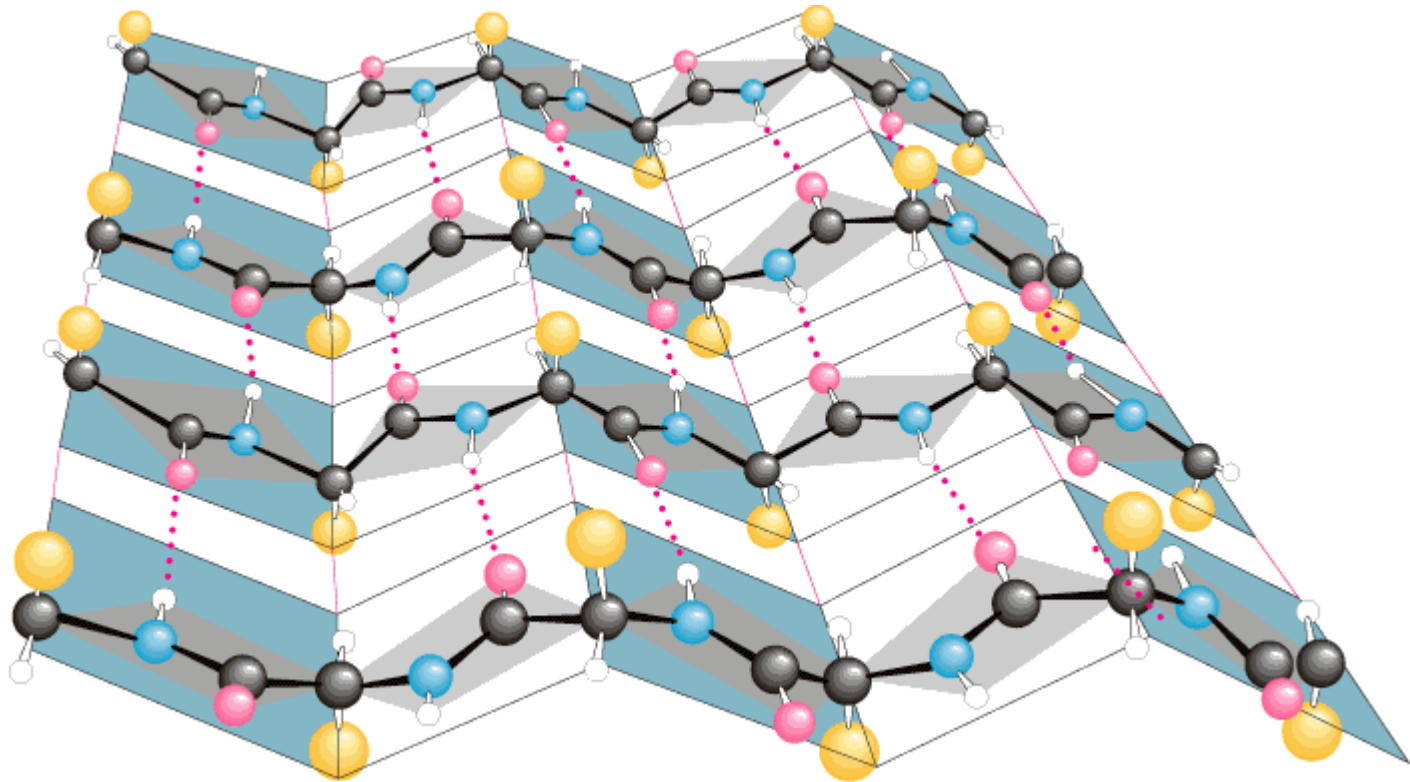
Hb {i.i+3}

Not parallel

# Proteine di membrana

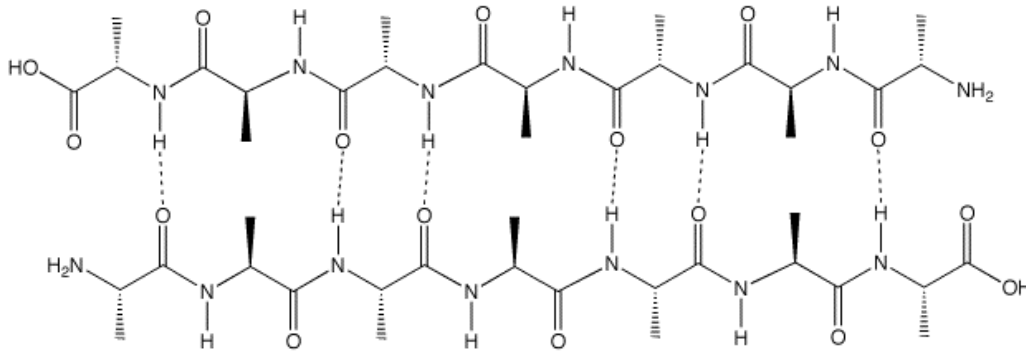


# Foglietti $\beta$

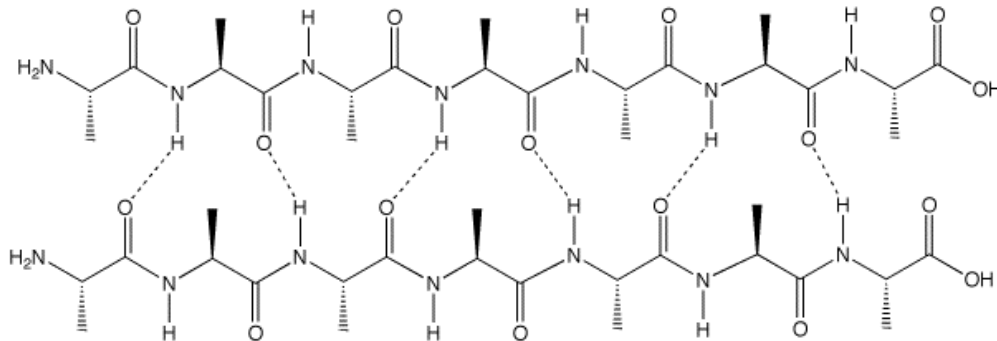


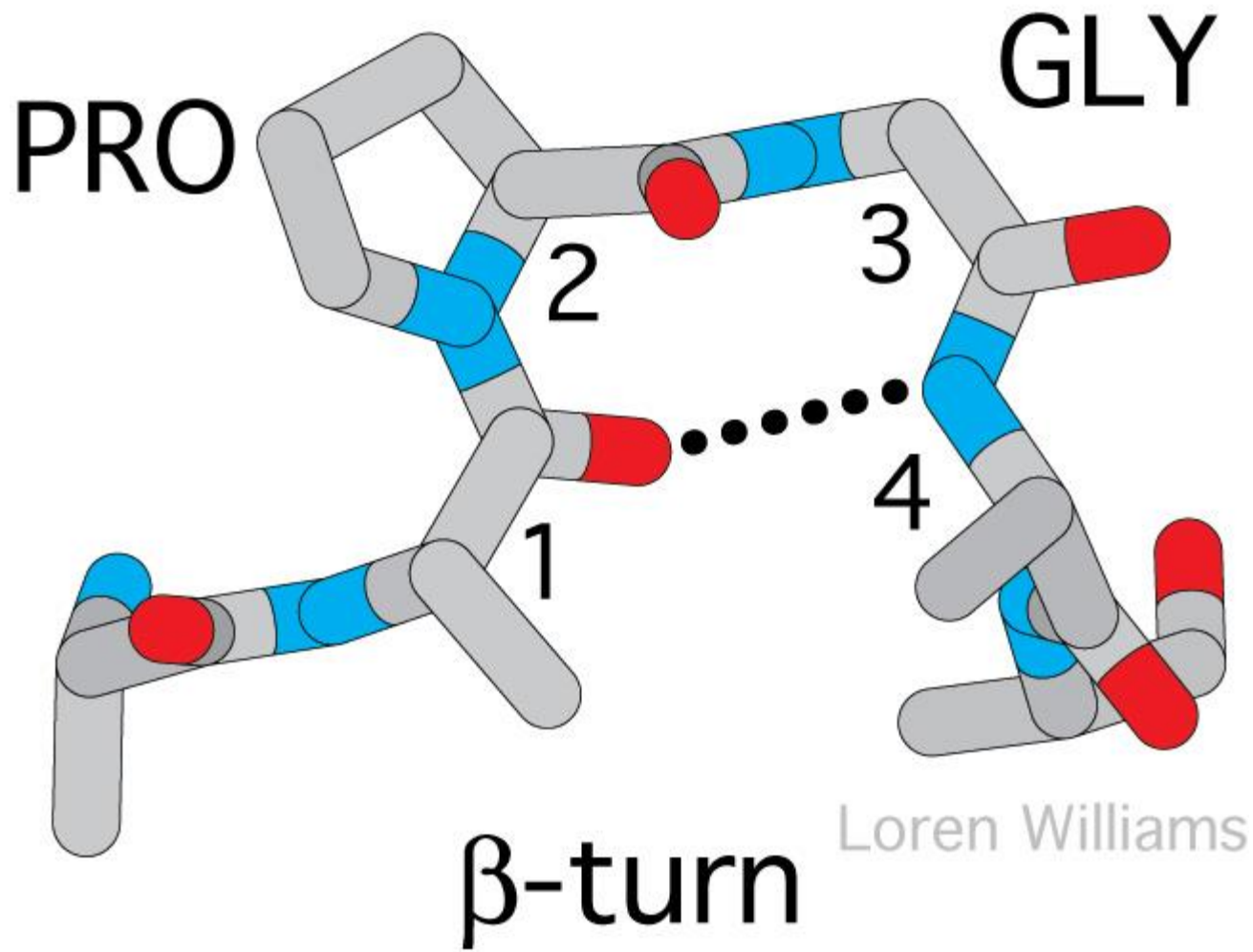
# Foglietti $\beta$

Anti-parallel

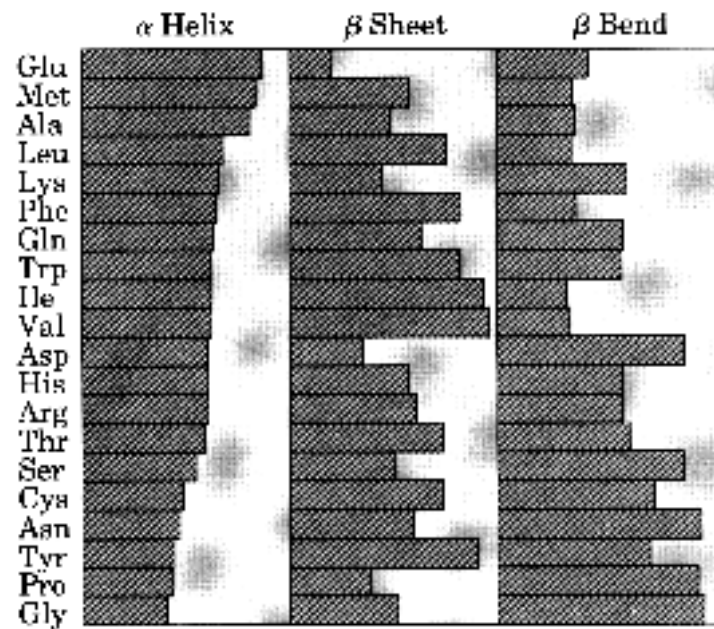


Parallel





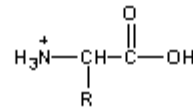
## Propensita' degli ammino acidi nelle strutture secondarie



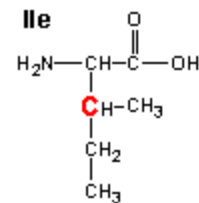
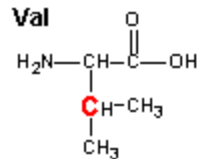
**Figure 7-12** Relative probabilities that a given amino acid will occur in the three common types of secondary structure.



## AMINO ACID PROPENSITIES FOR SECONDARY STRUCTURE



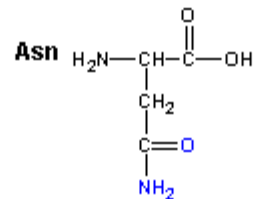
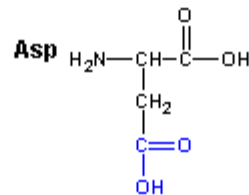
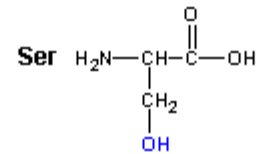
Branched at  $\text{C}_\beta$



Branch at  $\text{C}_\beta$   
**destabilizes  $\alpha$  helix**

**OK in  $\beta$  Sheet**  
side chain projects  
out of plan of main  
chain

No branch at  $\text{C}_\beta$  and  
R group can form H bonds

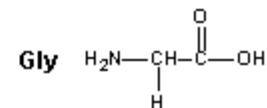


**destabilizes  $\alpha$  helix**  
H Bond donor/acceptors  
compete with main chain

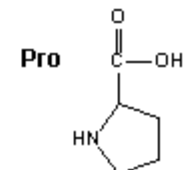
No branch at  $\text{C}_\beta$  and R  
group can't form H bonds

**OK in  $\alpha$  helix**  
side chain projects  
out of

Exceptions:



- too conformationally  
flexible

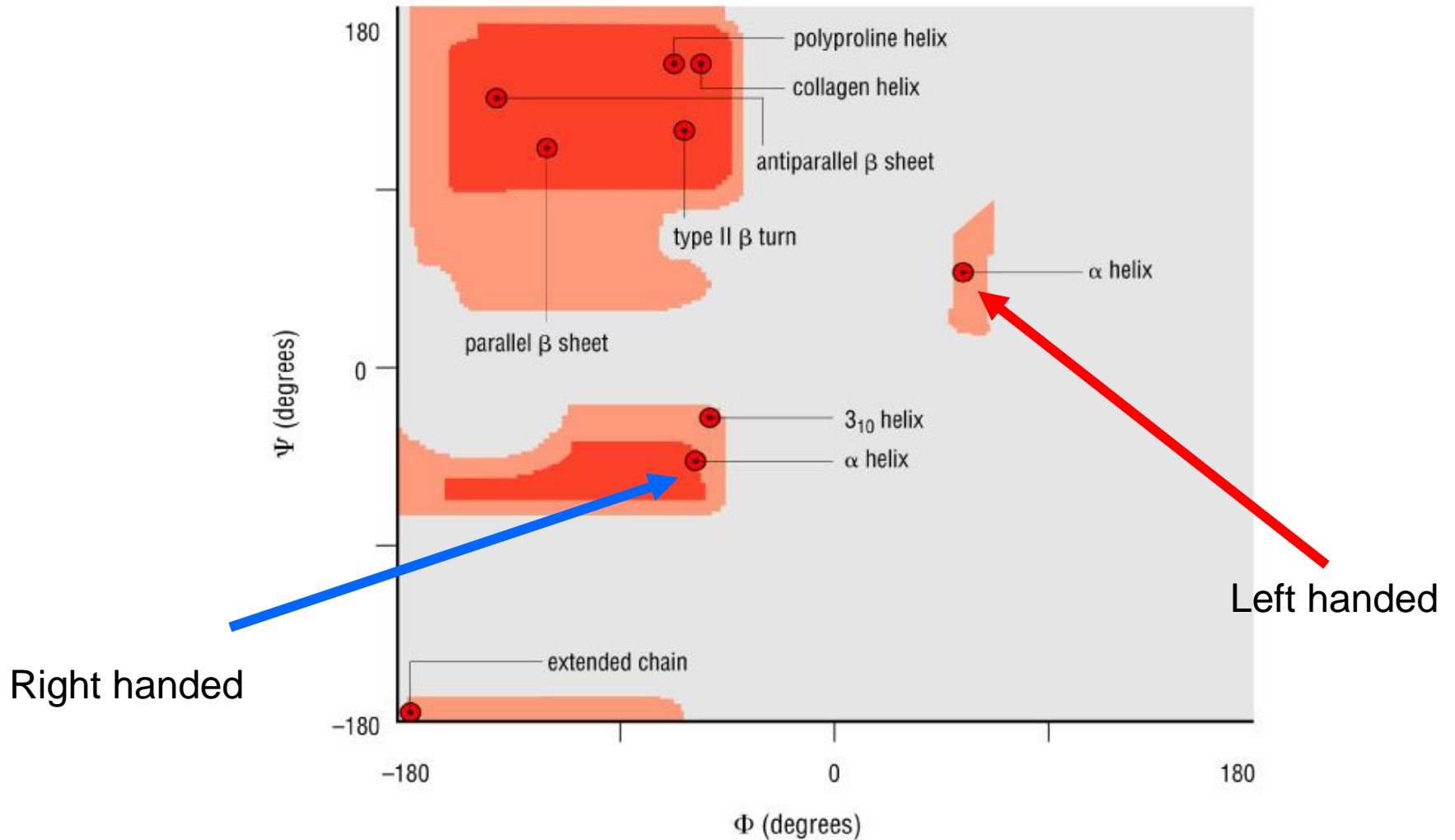


- too rigid

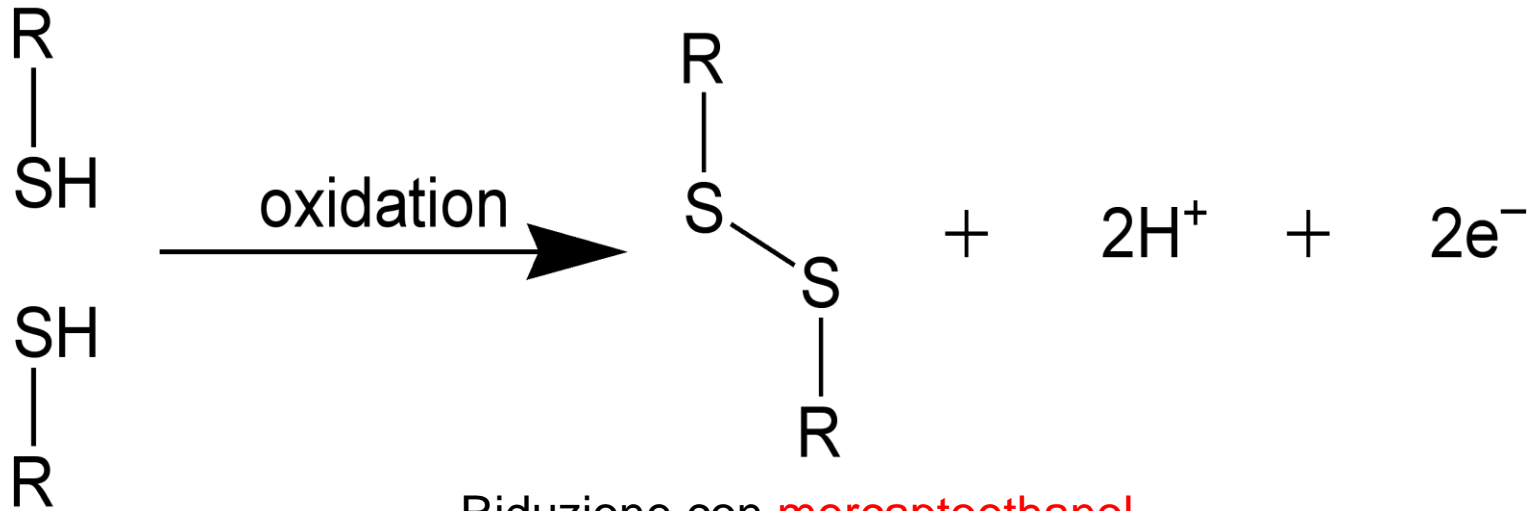
Ala and  
Leu

# Grafico di Ramachandran

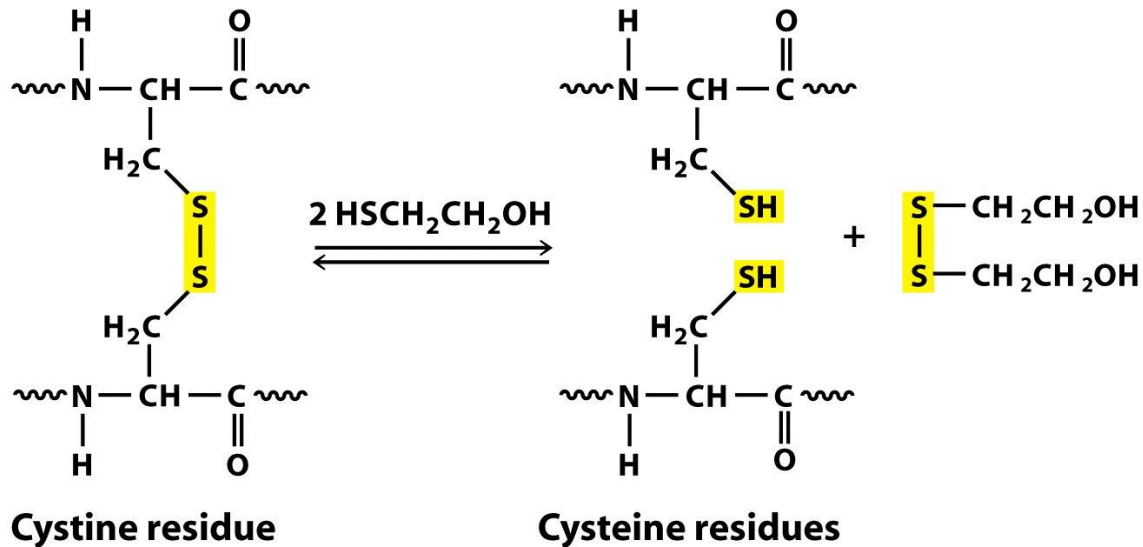
From [Protein Structure and Function 2004-2005 Online Update](#)  
by Gregory A Petsko and Dagmar Ringe



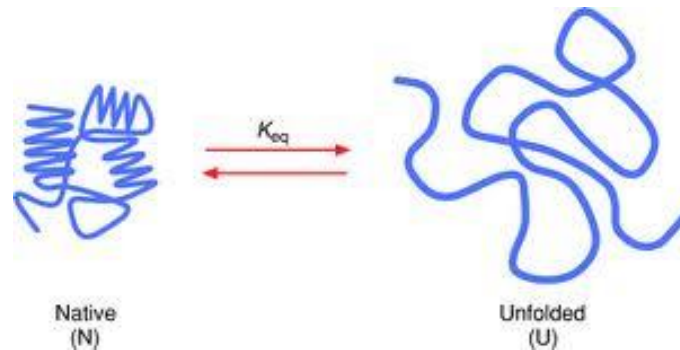
## Legami disolfuro



Riduzione con **mercaptoethanol**,



## Unfolding by denaturants



As substances such as urea, guanidinium and many alcohols are added to a solution, a protein is converted to a state that is qualitatively similar to the unfolded state induced by heating..

**A protein unfolded by denaturants as almost no residual structure**

Denaturants unfold proteins by interacting favourably with the protein interior

**For example urea can hydrogen bond with backbone amides and carbonyls of the peptide chain**

Buried residues of a protein have an unfavourable interaction with water.

**As the denaturant concentration increases, the unfavorable interaction with water is offset by an attractive interaction with the denaturant.**

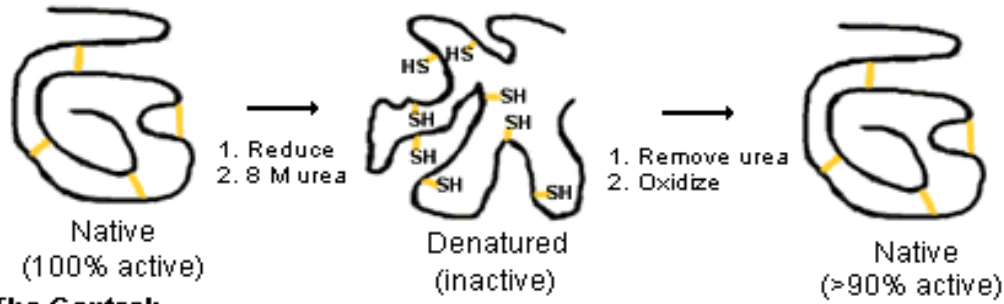
# Esperimento di Anfinsen (1960's)

His summary of the experiments was presented as a Nobel Prize Lecture and published in:  
Anfinsen, C.B. (1973) "Principles that govern the folding of protein chains." *Science* **181** 223-230.

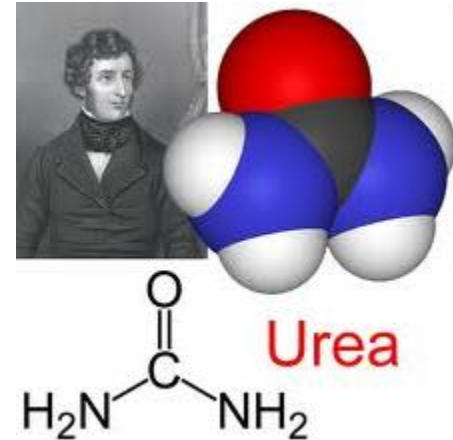
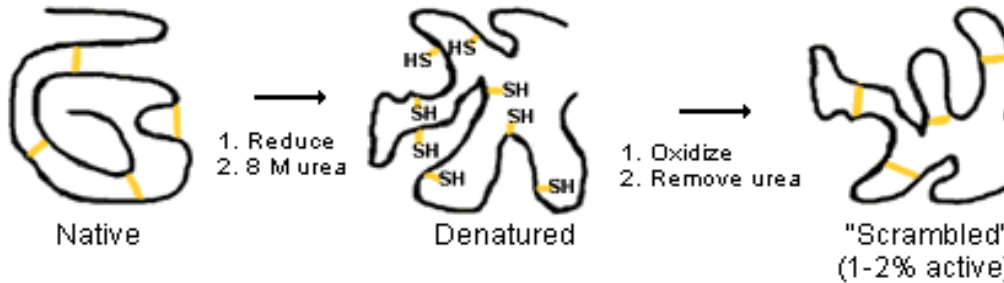


# Esperimento di Anfinsen (1960's)

## The Observation:



## The Control:

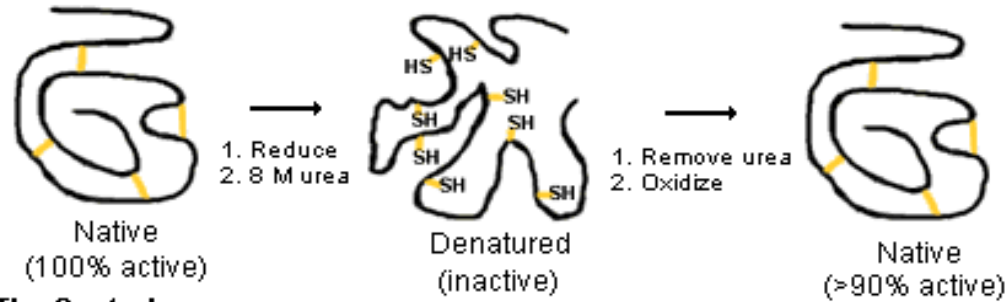


Ribonuclease A (RNaseA) is an extracellular enzyme of 124 residues with four disulfide bonds. In the first phase of the experiment, the S-S bonds were reduced to eight -SH groups (using mercaptoethanol, HS-CH<sub>2</sub>-CH<sub>2</sub>-OH); the protein was then denatured with 8 M urea. Under these conditions, the enzyme is inactive and becomes a flexible random polymer. In the second phase, the urea was slowly removed (dialysis); then the -SH groups were oxidized back to S-S bonds. If the protein was able to regain its native structure spontaneously after removal of the urea, we expect that it would also regain its activity. In fact, the activity was >90% of the untreated enzyme. Moreover, sequence analysis showed that nearly all of the correct S-S bonds had been formed.

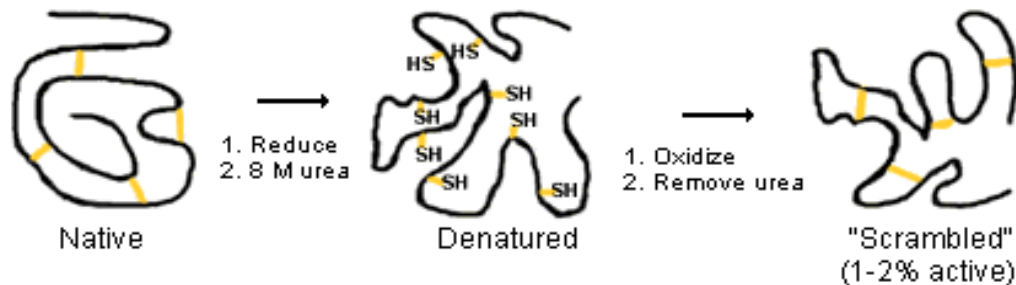
And if RNaseA was not completed unfolded???

# Esperimento di Anfinsen (1960's)

## The Observation:



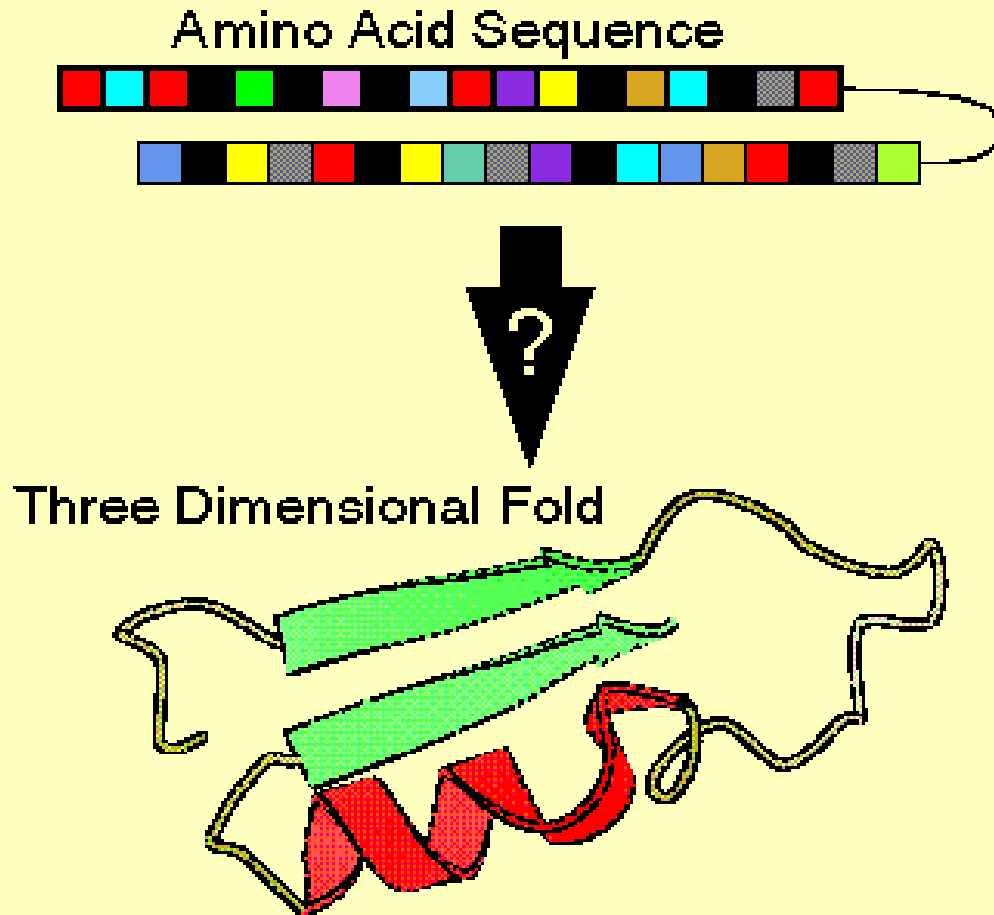
## The Control:



2

A reasonable objection can be raised to the above result by suggesting that perhaps RNaseA was not completely unfolded in 8 M urea. To address this class of objections, RNaseA was first reduced and denatured as above. But in the second phase, the enzyme was first oxidized to form S-S bonds, and **then** the urea was removed, *i.e.* the order of steps in the second phase of the experiment was reversed. The resulting activity was only about 1-2% of the untreated enzyme. Sequence analysis showed a random assortment of S-S bonds ("Scrambled" in the diagram). [Question: Can we account for the 1-2% recovery of activity in the "Scrambled" sample?].

# The Protein Folding Problem



**The native state is uniquely determined by the sequence**

- **The native state is thermodynamically stable and reachable from different starting conditions.**
- **Only few sequences are proteins**
- **Only few conformations are native states**
- **The folding time is very rapid (0.01-100 sec)**



## Contributions to the total free energy of the protein

1. Conformation entropy corresponding to the loss of degrees of freedom due to the bonding of amino acids and the restricted motion of side chains
2. Energy of intramolecular hydrogen bonds and of hydrogen bonds between the protein and external water molecules
3. Energy of van der Waals bonds
4. Coulomb energy of electrostatic bonds and coupling energy between dipoles formed by helices
5. Valence bond energy in disulphide bridges
6. Hydrophobic effect

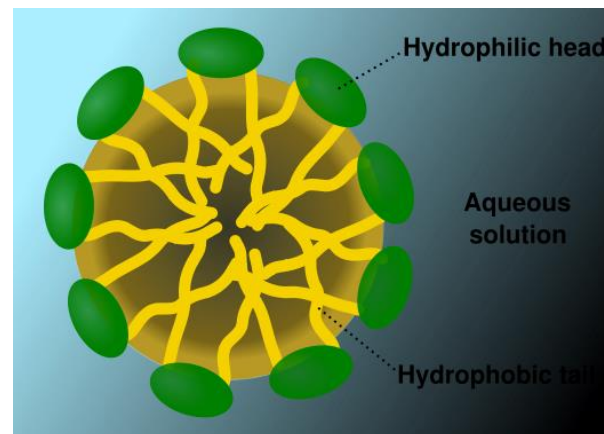
STABILITY:  $10kcal \cdot mol^{-1} = 20k_B T$  100 aa

# Effetto idrofobico

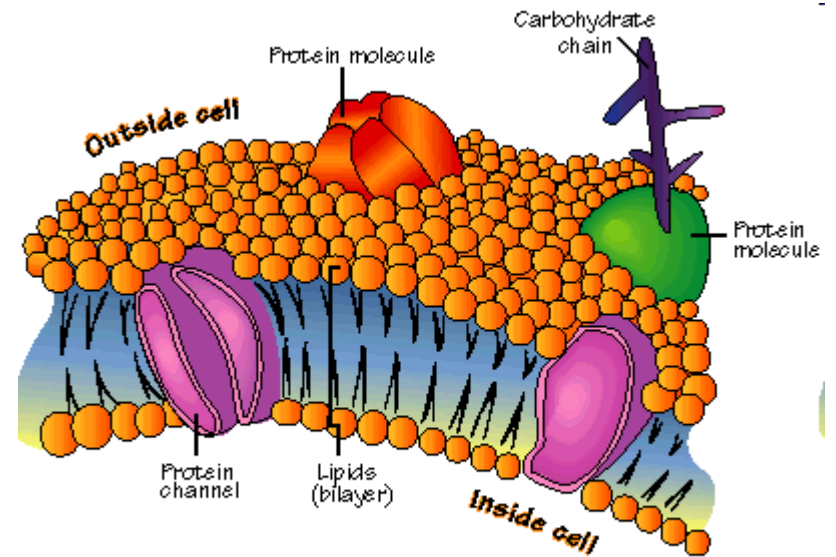
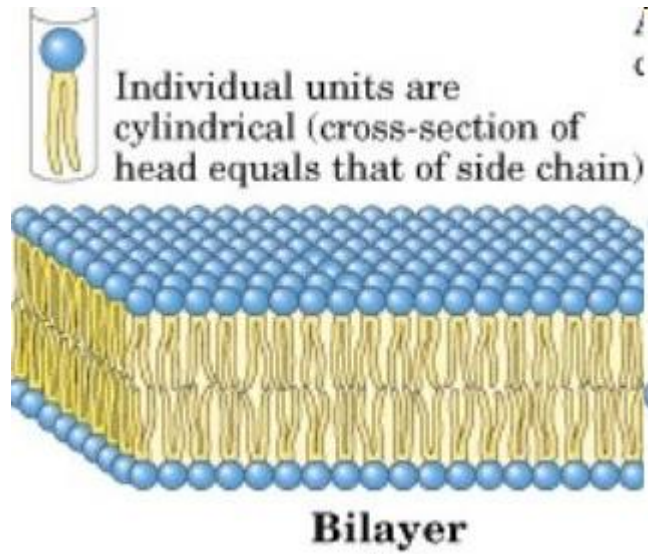
Le interazioni fra acqua e superfici non polari non sono favorevoli: proprio come l'olio disperso in acqua tende a raccogliersi in una unica goccia, anche i gruppi non polari nelle proteine tendono ad aggregarsi, per ridurre la superficie apolare a contatto con l'acqua.

Questa preferenza di **specie non polari** per ambienti non acquosi viene detto effetto idrofobico: esso e' uno dei principali fattori di stabilita' delle proteine.

L'effetto idrofobico fa si che sostanze non polari minimizzino il loro contatto con l'acqua, e molecole anfipatiche formino micelle in soluzioni acquose



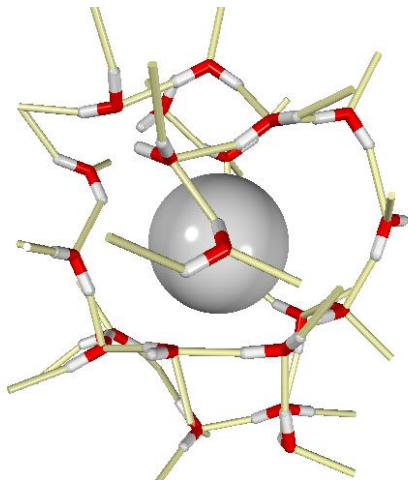
# Lipid bilayer



# Effetto idrofobico

Le molecole d'acqua allo stato liquido formano dinamicamente un alto numero di legami idrogeno.

L'introduzione di una molecola non polare nell'acqua, che temporaneamente rompe alcuni legami idrogeno fra le molecole d'acqua, poiche' un gruppo non polare non puo' ne' accettare ne' donare legami idrogeno con le molecole d'acqua



Le molecole d'acqua spostate si orientano per formare il maggior numero di nuovi legami idrogeno, creando una struttura ordinata, una specie di gabbia, detto clatrato, intorno alla molecola non polare

## Effetto idrofobico

Poiche' il numero di modi con cui le molecole d'acqua formano legami idrogeno sulla superficie di un gruppo non polare e' inferiore a quello che farebbero in sua assenza si ha una diminuzione di entropia del sistema.

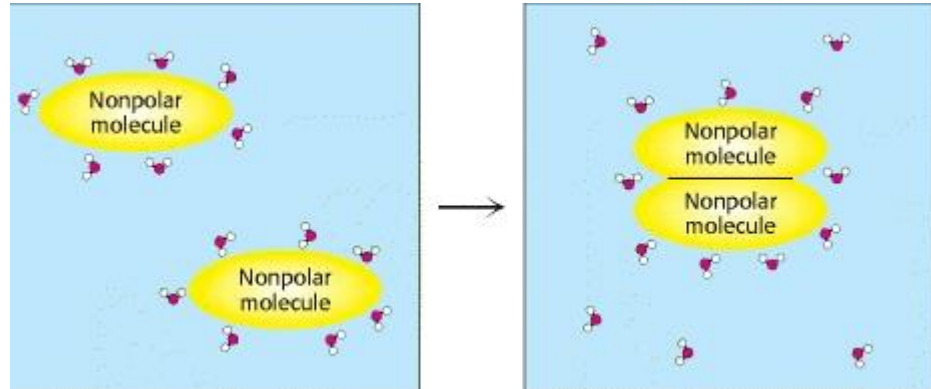
Anche se da un punto di vista entalpico il sistema clatrato e' piu' stabile

$$\Delta H < 0$$

per una debole liberazione di energia dovuto alla formazione di legami idrogeno ed interazioni di van der Waals, globalmente

$$\Delta G = \Delta H - T\Delta S > 0$$

# Effetto idrofobico

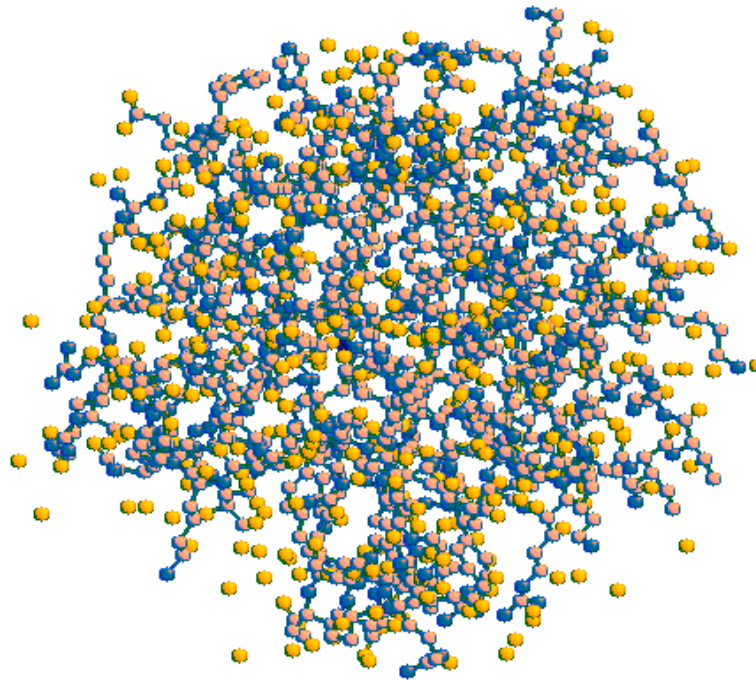


Perche' il processo sia spontaneo occorre l'aggregazione dei gruppi non polari in modo da minimizzare l'area superficiale della cavita' occupata dal gruppo apolare e quindi la perdita di entropia del sistema

**Effetto idrofobico e' favorito dall'aumento della temperatura**

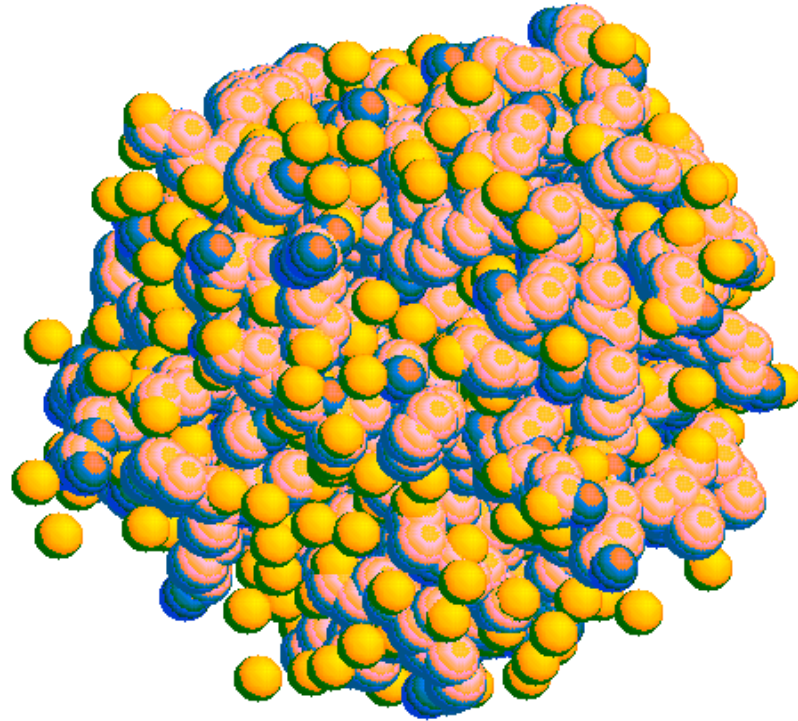
# Integrin Cell Adhesion Protein

184 Aminoacids 1491 Atoms



# Integrin Cell Adhesion Protein

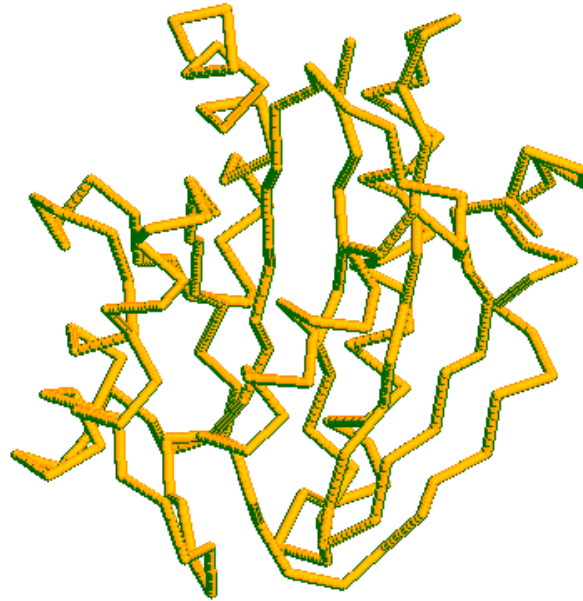
184 Aminoacids 1491 Atoms



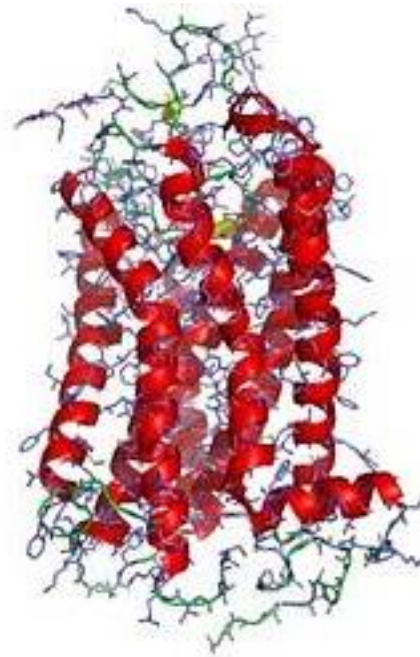
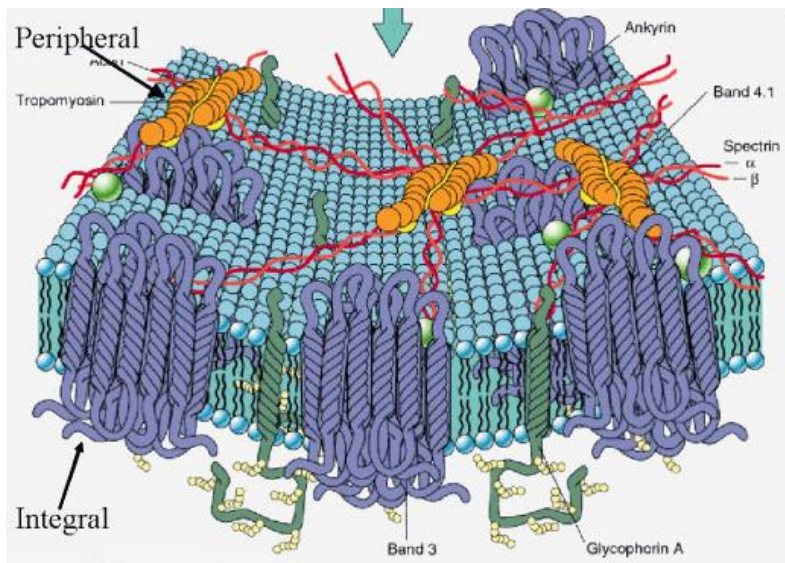


# Integrin Cell Adhesion Protein

184 Aminoacids 1491 Atoms



# Proteine di membrana



Only 1000 folds!!!!

# Growth Of Unique Folds (Topologies) Per Year As Defined By CATH (v4.0.0)

number of folds can be viewed by hovering mouse over the bar

